

## Web Tutorial

The ESPrict interface is composed by:

- A **BUTTONS** frame, fixed at the top of the page.
- A MAIN frame, containing the user form.
- A POP-UP window, containing the results of your ESPrict job.

### 1 The MAIN frame:

- Fill up the form by, at least, uploading a multiple alignment file in the section **Aligned Sequences**.
- PDB files and DSSP files can be uploaded in the section **Secondary structure depiction**.
- The rest of the form allows you to change parameters related to the secondary structures and sequence similarities depiction as well as the alignments output layout or the size and format of the resulting figures (PostScript, PDF, PNG, TIFF).
- All these options are detailed in the **User Guide** section or directly from the interface by clicking on the **?** icons.
- Short notices are also available for form items by hovering the pointer over a **i** icon.

### 2 The BUTTONS frame:



- Only yellow buttons are active at the exception of the **TIME** bar. Blue buttons are not clickable.
- When the main form is filled, click on the **SUBMIT** button to let ESPrict process your query.
- A result pop-up window automatically appears within seconds. This results window can be (re)opened at any moment by clicking on the **RESULTS** button.
  - In order to access the results, you may be required to **authorize your browser to display pop-up windows** from `esprict.ibcp.fr`. If needed, please refer to your browser documentation.
- The **DOC** button displays the full **User Guide** in a separated window.
- Click on **ADV** (ADVanced) or **EXP** (EXPerT) to have access to more options. The default mode is **BEG** (BEGinner).
- In **ADV** mode, you can introduce another secondary structure file, change labels of secondary elements, fiddle with special characters and calculate similarity scores between groups of sequences.
- In **EXP** mode, you can also define your own colors, shift sequence numbering, etc.
- You can navigate between **BEG**, **ADV** and **EXP** modes without losing information in your query.
- If you are in **ADV** or **EXP** mode, you can use the **+1** button to build a layered ESPrict figure. If you click on **+1**, a new layer is created (up to 10 layers can be defined). The parameters specified for **Layer 0** (i.e. the first one) are copied to **Layer 1** (and so on). Button **-1** allows the user to suppress the last created layer. Layer **Common** contains parameters shared in all layers.
- You can switch between the different layers by clicking on the tab bar (see below). Check **example #3** to learn more on this option.

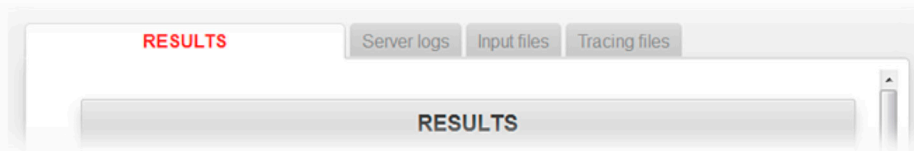


- Use the **SAVE** button to save your session on your computer, so that you can use later the same parameters and files.
- Use the **LOAD** button to load a previously saved session.
- Pay attention to the **TIME** bar. You must at least execute one command each 60 minutes otherwise your session will be closed.
- Before leaving, click on the **EXIT** button so the processing and results files are permanently removed from server.

### 3 The RESULTS pop-up window:

- Figures produced by ESPrict appear in this multi-tab window with dedicated links. Click to visualize them or click with the right button of the mouse for retrieve.

- Expert (or curious!) users can click on the `Server logs` `Input files` or `Tracing files` tabs to access the corresponding reports and data.



#### 4 EXAMPLE #1 - Beginner Mode

Session `vp7_beg.sav` for beginners (resulting PDF, PostScript, PNG).

Sequences homologous to `vp7_btv10`, a viral protein whose 3D structure has been solved, have been aligned with ClustalW. The form `vp7_beg.sav` allows the user to create a figure from the alignment and to display secondary structure elements of `vp7_btv10`. It has been created as follows:

- Start **ESPrpt**.
- Save the alignment file `vp7aln` on your disk, and upload it in the first section `Aligned Sequences` of the form.
- SUBMIT** and open the resulting image. A column is framed in blue if more than 70% of its residues are similar according to physico-chemical properties (threshold is set to 0.7 in the section `Sequence similarities depiction parameters` of the form and the button `%Equivalent` is activated). Note that `vp7_btv10` is the second sequence in the alignment.
- Save the PDB file `vp7_btv10.pdb` on your disk and upload it in the second section `Secondary structure depiction` of the form.
- SUBMIT** and open the resulting image. Observe that the secondary structure elements displayed above sequence blocks are named `vp7_btv1s`. This is normal: by default they are aligned with the first sequence displayed by ESPrpt, here `vp7_btv1s`. However, this is wrong: your secondary structure elements refer to `vp7_btv10`.
- Browse the form to the section `Defining groups`. Change the string `all` to `all`.
- SUBMIT** and open the resulting image. `vp7_btv10` is now the first displayed sequence and secondary structure elements are correctly aligned with it.
- Check the section `Alignments output layout`. You will see a yellow tooltip indicating "Suggested number of columns: 60 (for A4, portrait and a font size of 7)". This indicates the `Number of columns` to set in order to obtain a justified figure.
- SUBMIT** and **SAVE** session.

#### 5 EXAMPLE #2 - Advanced Mode

Session `vp7_adv.sav` for advanced users (resulting PDF, PostScript, PNG).

This session uses the same ClustalW alignment as in Example #1. You can start with the session `vp7_beg.sav` described above.

- Start **ESPrpt**.
- LOAD** previous session by using the buttons frame.
- SUBMIT** to check if everything is fine.
- Click on **ADV** to access new options.
- Check the button `Relative accessibility` in section `Secondary structure depiction` of the form, to show accessibility per residue with a blue bar below sequence blocks on the image.
- Click on `Hide names` in the same section, to remove the string `vp7_btv10` above sequence blocks, at the same line as secondary structure elements.
- SUBMIT** to check.
- Type the command `U R 127 250` in section `Special commands and characters`, to add two red triangles below residues 127 and 250 of the first displayed sequence, `vp7_btv10`. These triangles show the separation between an  $\alpha$ -domain (1-127, 250-349) and a  $\beta$ -domain (128-249) in the 3D structure.
- SUBMIT** to check.
- In the same section, type `S B 168-170 178-180` to mark an RGD tripeptide observed at different positions in sequences. Then type:
  - `X B 1-126 254-349` to color secondary structure elements of the  $\alpha$ -domain in blue.
  - `X G 127-253` to color secondary structure elements of the  $\beta$ -domain in green.
  - `T R 1 2` to color in red the names of the first and second displayed sequences.
  - Note:** you must press [ENTER key] after each line of special commands.
- SUBMIT** to check.
- Type in subsection `Footnotes` of `Alignments output layout` the sentence: `Alignment for protein vp7`.
- SUBMIT** to check.
- In section `Defining groups`, delete `2 all` and type `2 1 3-6 7-8 [ENTER key] 9` in order to define three groups of sequences according to phylogeny (the first group is made of `btv` sequences only) and to calculate in-group as well as cross-group similarities. This necessitates the use of a scoring matrix, and the option `Risler` is selected instead of `%Equivalent` in section `Sequence similarities depiction parameters`.

- **SUBMIT** to check. You can now observe:
  - At column 1 that all residues are identical and are boxed in red.
  - At column 10 that residues of the first group are identical and are in red (all threonines) and that those of the second are similar (threonine and serine) and are in red ; the third group is made of a single sequence and the residue is in black. The global similarity score calculated from all groups is > 0.7 (which is the threshold entered in the section `Similarity Calculation` ) and the column is framed in blue.
  - At column 19 that residues are not framed, the global similarity score being < 0.7. Moreover, residues of the second group are in black, because they are not similar (lysine and threonine) according to a in-groups score < 0.7 (the same threshold is used by the program for global scores and in-groups scores).
  - At column 79 that the column has a yellow background. Residues are similar in each group (in-groups scores > 0.7) but significantly different from the first group (all prolines) to the second (all histidines). Thus, the cross-groups score is > 0.5, which is the current threshold according to the section `Sequence similarities depiction parameters` .
- **Remark:** try this if you want a more colorful figure:
  - Select option `Thermal` in section `Alignments output layout` to obtain bold characters.
  - Type the command `M Y a11` in section `Special commands and characters` to add a yellow background on similar residues.
  - **SUBMIT** to check.

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EXAMPLE #3 - **Expert Mode**

Session `vp7_exp.sav` for experts (resulting [PDF](#), [PostScript](#), [PNG](#)).

Again, same CLustalW alignment, but information on the 3D structure of `vp7_btv1s` is now entered. The button **+1** of the buttons frame is used for this purpose. The following must be generated in **ADV** or **EXP** mode. We will start from scratch.

- Open **ESPrpt**.
  - Upload `vp7.a1n` in the section `Aligned Sequences` .
  - Upload `vp7_btv10.pdb` in the section `Secondary structure depiction` .
  - Type `X B 1` in the section `Special commands and characters` , to color in blue secondary structure elements and align them with the first displayed sequence.
  - Type `h` in subsection `Replace secondary structures labels` of `Special commands and characters` , in order to remove the label `h1` from the `310`-helix.
  - Change `a11` to `2 a11` in the section `Defining groups` to display `vp7_btv10` as first sequence in the alignment.
  - **SUBMIT** to check the result.
  - Click on `Hide sequences` in the first section and **SUBMIT** again. Only secondary structures elements are now displayed.
  - Click on **+1** in the buttons frame to create a new layer.
  - The form now contains two independant layers named `Layer 0` and `Layer 1` and a shared one ( `Common` ). Check that the alignment file `vp7.a1n` has been copied from `Layer 0` to `Layer 1` .
- All following commands must be entered in `Layer 1` .
- Disable option `Hide sequences` , then enter a `Vertical shift` of -1 in section `Alignments output layout` .
  - **SUBMIT** to check. You now have a gap between secondary structure elements of `vp7_btv10` and sequences.
  - Enter `vp7_btv1.dssp` in the section `TOP secondary structures` of the box `Secondary structure depiction` , which corresponds to `vp7_btv1s`.
  - Type `X R 2` in the section `Special commands and characters` , to color in red secondary structure elements and align them with the second displayed sequence.
  - Type `2 a11` in the section `Defining groups` .
  - **SUBMIT** to check.
- Information on secondary structure elements are well aligned with sequences but labels of `vp7_btv1s` need to be removed.
- Click on `Hide labels` in section `Secondary Structure` .
  - **SUBMIT** to check.
- All information is now well displayed. We will add extra boxing on residues.
- Type `Q P 1-3 169-171` in section `Special commands and characters` . This means that columns 169-171 of sequences 1-3 are boxed in pink. Note that the program uses columns numbering instead of residues numbering for the special commands `Q`, `V` and `W`. You can visualize columns numbers by checking the button `Ruler` in the same section.
  - Type `Q P 5-7 169-171 [ENTER key] Q P 8 180-182` in the same section to box all `RGD` tripeptides.

- Select the mode `Flashy` in section `Color scheme` of the `Common` tab to add a yellow background on similar residues.
- **SUBMIT** to check.

→ We will now color sequence names of `vp7_btv10` and `vp7_btv1s`. Type in the same section `Special command and characters`.

- `T B 1` to color the name of the first displayed sequence in blue.
- `T R 2` to color the name of the second displayed sequence in red.

→ Finally, we will number all sequences.

- Click on `Number sequences` in section `Aligned Sequences` of `Layer 0`.
- **SUBMIT** to check.

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The three examples above refer to figures prepared in the group of Prof. David STUART, [Division of Structural Biology](#), Oxford.